



# **IGF-1 ELISA Kit Instructions**

For the quantitative determination of IGF-1 in  
human serum, plasma, and cell culture media

**Catalog #80572  
96 Assays**

**For research use only. Not for use in diagnostic procedures.**

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**A. Intended Use**

The IGF-1 ELISA kit is for the quantitative determination of IGF-1 in human serum, plasma, and cell culture media. Please read the complete kit insert before performing this assay. The kit is for RESEARCH USE ONLY. It is not intended for use in diagnostic procedures.

**B. Introduction**

IGF-1 (Insulin-like Growth Factor-1) plays a critical role in regulating the proliferation and differentiation of many cell types. IGF-1's major regulators are growth hormone and nutrition.

**C. Principle of the Assay**

The IGF-1 ELISA kit is an ELISA sandwich assay for human IGF-1. It utilizes two specific and high affinity antibodies for this protein. IGF-1 is first dissociated from the IGF-BPs via an acidic buffer. After neutralization of the samples, the excess IGF-2 occupies the IGF-binding sites of the binding proteins, thus allowing the measurement of resulting free IGF-1. The IGF-1 in the human sample binds to the first antibody coated on the microtiter plate. In the following step, the biotinylated and Streptavidin-Peroxidase conjugated second specific anti-IGF-1 antibody binds in turn to the immobilized IGF-1. In the closing substrate reaction, the IGF-1 levels of the samples can be measured by color intensity.

**D. Kit Storage**

1. Upon receipt of the IGF-1 ELISA kit, store it at 2-8°C and avoid light exposure (do not freeze the kit or hold it at temperatures above 25°C).
2. The kit should not be used after the expiration date.

**E. Assay Materials**

**E.1. Materials provided**

**TABLE 1 Contents of the kit**

Mark	Description	Amount
MIC	Antibody-coated Microplate (12 x 8)	1 pack
STD1-5	Standards	1 x 5 vials
DIL	Dilution Buffer	1 x 25 mL
CON1-2	Controls	1 x 2 vials
AB CONJ	Antibody Conjugate	1 x 9 mL
ENZ CONJ	Enzyme Conjugate	1 x 12 mL
WASH	Wash Buffer (20X Concentrate)	1 x 50 mL
SUB	Substrate Solution	1 x 12 mL
STOP	Stop Solution	1 x 12 mL
	Sealing tape for plate	2x, adhesive

**E.2. Materials required but not provided**

Micropipettes and disposable tips  
Distilled or deionized water  
Polypropylene microtubes  
Vortex mixer  
Microplate shaker (350 rpm)  
Microplate reader (capable of reading  $A_{450}$  and  $A_{630}$  values)

**F. Assay Precautions**

1. Only appropriately-trained personnel should use the kit. Laboratory personnel should wear suitable protective clothing. All chemicals and reagents should be considered potentially hazardous. Avoid ingestion and contact with skin and eyes.
2. Some assay components may contain human sourced materials. Accordingly, all assay components should be handled as if potentially infectious using safe laboratory procedures.
3. Do not use the reagents after the expiration date.
4. Reagents are light sensitive and should be protected from sunlight.

**G. Maximizing Kit Performance**

1. Given the small sample volumes required (10  $\mu$ L), pipetting should be done as carefully as possible. A high quality 20  $\mu$ L or better precision pipette should be used for such volumes. Drops of liquid adhering to the outside of the pipette tips should be removed by wiping to ensure the highest degree of accuracy.
2. In order to prevent the microplate wells from drying out and to get the best results, samples and reagents should be dispensed quickly into the wells.
3. Each standard and sample should be assayed in duplicate.
4. The same sequence of pipetting and other operations should be maintained in all procedures.
5. Do not mix reagents that have different lot numbers.

**H. Sample Collection**

Serum and plasma samples collected with EDTA, Citrate, or Heparin anticoagulant can be used. Cell culture media can also be used as a sample matrix. Undiluted samples should be chilled as soon as possible after sample withdrawal. For longer term storage, samples can be stored at  $-20^{\circ}\text{C}$ . Avoid repeated freeze-thaw cycles of samples.

**I. Assay Procedure**

**I.1. Preparation of reagents**

1. Antibody-coated microplate  
Provided as ready to use. Protect from moisture.
2. Standards 1-5  
Standards are provided in lyophilized form with concentrations ranging from 2 ng/mL to 50 ng/mL. Dilute each standard with 0.5 mL of Dilution Buffer (marked "DIL"). After reconstitution, it is recommended that standards be allowed to sit for 15 mins at room temperature and then mixed thoroughly but gently with a Vortex mixer. Reconstituted standards are stable for three months at  $-20^{\circ}\text{C}$ . Standards should not be repeatedly thawed (at most, three times), so standards should be appropriately aliquoted in appropriate volumes prior to being frozen. Standards are provided in the following concentrations: 2, 5, 15, 30, and 50 ng/mL.

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3. Dilution Buffer  
Provided as ready to use.
4. Controls 1-2  
Controls are provided in lyophilized form with target value and ranges included on their labels. Reconstitute controls with 0.5 mL of Dilution Buffer. Reconstituted controls are stable for three months at -20°C. Reconstituted controls should not be repeatedly thawed (at most, three times).
5. Antibody Conjugate  
Provided as ready to use.
6. Enzyme Conjugate  
Provided as ready to use.
7. Wash Buffer (20X Concentrated)  
The wash buffer has to be diluted 1:20 with distilled or deionized water prior to use. For example, 50 mL of wash buffer must be diluted with 950 mL of distilled or deionized water. Wash buffer is stable for 4 weeks at 2-8°C after dilution, so dilute only as needed.
8. Substrate Solution  
Provided as ready to use.
9. Stop Solution  
Provided as ready to use.

### I.2. Dilution of samples and controls

1. Samples and controls need to be diluted with Dilution Buffer for use with the assay. Please note that this section only applies to samples and controls, not standards. Samples should be diluted at least 1:10 in dilution buffer. A sample dilution of 1:21 using 10 µL of sample is generally suitable for serum and plasma samples. Allow diluted samples to incubate at room temperature for at least 15 minutes. Reconstituted controls should be diluted in the same ratio as the samples. Samples and controls must be used within 2 hours once diluted.

Since IGF-1 levels can vary, dilution ratio may need to be adjusted as appropriate, particularly for cell culture samples.

### I.3. Assay procedure

Prior to running the assay, all reagents should be brought to room temperature for at least 30 minutes. Reagents should be stored at 2-8°C immediately after use. Before use, mix the reagents thoroughly by gentle agitation or swirling.

1. Add 80 µL of Antibody Conjugate in all wells to be used.
2. In each well, add 20 µL of diluted sample or 20 µL of standard or 20 µL of diluted control and mix well by repeated pipetting.  
**Note:** A blank using 20 µL of Dilution Buffer is recommended.
3. Cover the wells with sealing tape and incubate the plate for 1 hour at room temperature (shake at 350 rpm).
4. Aspirate well contents and wash five times using 300 µL of Wash Buffer per well. After each wash, remove any remaining solution by inverting and tapping the plate firmly on a clean paper towel.
5. Add 100 µL of the Enzyme Conjugate in each well.
6. Cover the wells with sealing tape and incubate the plate for 30 minutes at room temperature (shake at 350 rpm).
7. Aspirate well contents and wash five times using 300 µL of Wash Buffer per well. After each wash, remove any remaining solution by inverting and tapping the plate firmly on a clean paper towel.
8. Add 100 µL of Substrate Solution in each well.

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9. Incubate the plate for 15 mins in dark room at room temperature.
10. Stop the reaction by adding 100  $\mu$ L of Stop Solution.
11. Measure absorbance within 30 minutes using a plate reader (measure  $A_{450}$  values and subtract  $A_{630}$  values).

### I.4. Determining the IGF-1 concentration

1. Using computer software, construct the IGF-1 calibration curve by plotting the mean change in absorbance value for each calibrator (incl. blank) on the Y axis versus the corresponding IGF-1 concentration on the X axis. A higher-grade polynomial, or four parametric logistic (4-PL) curve fit or non-linear regression are suitable for the evaluation.

**Note:** A calibration curve should be plotted every time the assay is performed.

2. IGF-1 concentrations in the samples are interpolated using the calibration curve and mean absorbance values for each sample. For diluted samples and controls, the values obtained must be multiplied by the dilution factor to obtain the final IGF-1 concentration. The IGF-1 concentration is expressed in ng/mL.

**Note:** Samples with high IGF-1 concentrations (ie. fall above the range of the assay) should be further diluted with the dilution buffer and rerun.

### J. Performance characteristics

#### J.1. Assay range

The IGF-1 ELISA Kit has an assay range from 2 – 50 ng/mL.

#### J.2. Precision

The assay has a within-run and total precision of CV < 10%.

### Warranty

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